

Journal of Pharmaceutical Advanced Research**(An International Multidisciplinary Peer Review Open Access monthly Journal)**Available online at: www.jparonline.comR
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3**Medicinal plants with antioxidant activity – A review****Mahdi M. Thuwaini**

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ABSTRACT:

The production of free radicals occurs continuously in all cells as part of normal cellular function. However, excessive production of free radicals can play a role in many diseases. Antioxidants prevent tissue damage caused by free radicals by inhibition of their production, scavenging them, or by promoting their decomposition. This review highlighted the natural sources of antioxidants to be used as beneficial supplements in prevention of many diseases.

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The body is constantly producing free radicals due to the regular use of oxygen. Oxidation is a chemical reaction that transfers electrons from one molecule to an oxidizing agent. Oxidation reactions are known to produce free radicals. These free radicals are responsible for the cell damage in the body and contribute to various kinds of health problems, such as heart disease, diabetes, macular degeneration, and cancer. Antioxidants are a class of chemical substances naturally found in our food that can prevent or reduce the oxidative stress of the physiological system. Plants are the major source of natural antioxidants. Many previous reviews revealed that medicinal plants are efficient sources of antioxidant and free radical scavenging activities ^[1-3].

Keywords: Medicinal plants, Antioxidant, Free radicals, Therapeutic.

Table 1. Medicinal plants with antioxidant effects.

Medicinal plant	Active extracts or compounds	Methods	Ref.
<i>Achillea santolina</i>	Hydroalcoholic extract	Lipid peroxidation inhibition assay and DPPH assay	7-9
<i>Adiantum capillus-veneris</i>	Leaf extract	Lipid peroxidation inhibition assay, ABTS and DPPH assays	10-13
<i>Adonis aestivalis</i>	Flavonoids, crude extract	ABTS and DPPH assays	14-16
<i>Agrimonia eupatoria</i>	Different extracts and fractions	ABTS and DPPH assays	17-18
<i>Ailanthus altissima</i>	Ethyl acetate (EtOAc) fraction	ABTS and DPPH assays	19-20
<i>Alhagi maurorum</i>	Aqueous extract	FRAP and DPPH assays	21-22
<i>Allium cepa</i>	aqueous extract	In vivo improve superoxide dismutase, catalase, glutathionperoxidase, and reduced glutathione	23
<i>Allium sativum</i>		DPPH, and inhibition of lipid peroxidation assays	24-25
<i>Alpinia galangal</i>	Extracts and essential oil	ABTS and DPPH assays and inhibition of lipid peroxidation assays	26-28
<i>Althaea officinalis</i>	Scopoletin (7-hydroxy-6-methoxy coumarin)	Inhibition of lipid peroxidation assay	29-30
<i>Ammanniabaccifera</i>	Methanolic extract	Inhibition of lipid peroxidation assay	31-32
<i>Ammi visnaga</i>	Butanolic extract	DPPH assay	33-34
<i>Anchusa italica</i>	butamol extract of <i>Anchusa italica</i> and two of the triterpenes' compounds	DPPH assay	35-36
<i>Anthemis nobilis</i>	Chamazulene, aqueous extracts	ABTS and Inhibition of lipid peroxidation assays	37-39
<i>Antirrhinum majus</i>	Absolute methanol extract	DPPH assay	40-41
<i>Apium graveolens</i>	leaf extract	NO, DPPH, ABTS, LPO, and HPO methods	42-43
<i>Arachis hypogaea</i>	Peanut peptide	DPPH assay	44-46
<i>Arctium lappa</i>	caffeoylquinic acid derivatives from the root	The antioxidant activities were measured in a hexane/2-propanol solution of methyl linoleate in the presence of a radical initiator	47-49
<i>Artemisia campestris</i>	Aqueous, Ethyl acetate extracts and essential oils	ABTS and DPPH assays and inhibition of lipid peroxidation assays	50-53
<i>Asparagus officinalis</i>	Juice, anthocyanins A1 and A2	Inhibition of lipid peroxidation and DPPH assay	54-55
<i>Astragalus hamosus</i>	Rhamnocitrin 4'- β -D-galactopyranoside (RGP) obtained from leaves	<i>In vivo</i> antioxidant enzymes	56-57
<i>Avena sativa</i>	Phenolic-rich fractions of oats and 3 avenanthramides compounds	DPPH assay, inhibition of lipid peroxidation	58-60
<i>Bacopa monniera</i>	Aqueous and alcoholic extract	DPPH assay, inhibition of lipid peroxidation and <i>in vivo</i> antioxidant enzymes	61-63
<i>Ballota nigra</i>	Aerial parts extracts, diterpenoid and flavonoid	DPPH assay and inhibition of lipid peroxidation	64-66
<i>Bauhinia variegata</i>	crude extracts and fractions	DPPH assay	67-68
<i>Bellis perennis</i>	The the aerial parts	DPPH assay	69-71
<i>Benincasa hispida</i>	Seeds and fruit extracts	DPPH, ABTS and <i>in vivo</i> inhibition of lipid peroxidation	72-75
<i>Bidens tripartita</i>	Crude extracts of the herb and flowers	DPPH assay	76-77
<i>Brassica nigra</i>	Crude extract	DPPH assay	78-79
<i>Brassica rapa</i>	Root extract and the carbohydrate derivatives of the roots	Inhibition of lipid peroxidation and DPPH assay	80-81
<i>Bryophyllum calycinum</i>	leaf extracts	DPPH assay	82-83
<i>Caesalpinia crista</i>	Seeds extracts	Inhibition of lipid peroxidation and DPPH assay	84-86
<i>Calamintha graveolens</i>	Crude extracts of the leaves and flavanone glycosides	DPPH and inhibition of lipid peroxidation	87-88
<i>Calendula officinalis</i>	Crude extracts	DPPH, ABTS and <i>in vivo</i> inhibition of lipid peroxidation and effect on antioxidant enzymes	89-91

<i>Calotropaprocera</i>	Crude extracts, latex, flavonoids and phenols	DPPH and inhibition of lipid peroxidation	92-94
<i>Canna indica</i>	The aerial parts methanolic extract	DPPH assay	95-96
<i>Capparis spinosa</i>	Aerial part and root extracts	DPPH, ABTS, FRAP assays	97-99
<i>Capsicum species</i>	Crude extracts and flavonoids	DPPH, ABTS, FRAP assays and α -carotene-linoleic acid system	100-103
<i>Carthamus tinctorius</i>	Seeds extracts, serotonin and flavonoids derivatives	DPPH, FRAP assays and inhibition of lipid peroxidation	104-107
<i>Carum Carvi</i>	Seeds extracts and essential oils	DPPH assay and inhibition of lipid peroxidation,	108-111
<i>Cassia occidentalis</i>	The methanolic extracts of leaves, stems and seeds	DPPH assay, nitric oxide scavenging activity, β -carotene-linoleic acid model system, hydroxyl radical scavenging activity, reducing power, metal chelating activity and superoxide radical scavenging activity	112-114
<i>Casuarina equisetifolia</i>	Crude extracts	DPPH and FRAP assays	115-116
<i>Celosia cristata</i>	Ethanol extract	DPPH, ABTS and FRAP assays	117-119
<i>Centaurea cyanus</i>	Crude extract	Chemiluminescence's method – system luminol/H ₂ O ₂	120-121
<i>Chenopodium album</i>	Ethanol leaf extract	DPPH assay	122-124
<i>Chrozophora tinctoria</i>	Methanol leaf extract	DPPH assay	125-126
<i>Cicer arietinum</i>	Crude seeds and roots extracts and lectin	DPPH and hydrogen peroxide radical, and <i>in vivo</i> effect on superoxide dismutase, catalase, GSH, increased MDA levels.	127-134
<i>Cichorium intybus</i>	Crude extracts and fractions,	(DPPH) and hydrogen peroxide radical, and <i>in vivo</i> effect on superoxide dismutase, catalase, GSH, increased MDA levels.	135-138
<i>Citrullus colocynthis</i>	Crude seeds extracts and flavonoids, isosaponarin, isovitexin and isoorientin 3'-O-methyl ether, isolated from the fruits	DPPH assay	139-144
<i>Citrus species</i>	The leaf extracts and leaf essential oil of <i>Citrus aurantifolia</i> , <i>Citrus medica</i> , <i>Citrus limonum</i> and <i>Citrus sinensis</i>	DPPH assay and inhibition of lipid peroxidation	145-153
<i>Clerodendrominerme</i>	Methanolic extract of the aerial parts, and compound -hydroxy-6,7,4'-trimethoxy flavones	DPPH Assay, reducing power assay and total antioxidant activity	154-156
<i>Clitoriaternatea</i>	Different solvent extracts of different parts	DPPH Assay	157-161
<i>Colchicum candidum</i>	Methanol and ethanol extracts	DPPH, ABST and FRAP assays	162-163
<i>Convolvulus arvens</i>	Crude extracts of the aerial parts	DPPH, ABST and FRAP assays	164-170
<i>Convolvulus scammonia</i>	The crude methanolic extract	DPPH Assay	171-172
<i>Cordia myxa</i>	Crude extracts	DPPH assay	173-175
<i>Coriandrum sativum</i>	Seed powder, essential oil and crude extracts	DPPH assay, lipoxigenase inhibition, phospholipid peroxidation inhibition, iron chelating activity, hydroxyl radical scavenging activity, superoxide dismutation, glutathione reduction and antilipid peroxidation	176-183
<i>Cotoneaster racimiflora</i>	Racemiside, scopoletin, 7,8-dimethoxy-6-hydroxycoumarin, 3,3',4'-tri-O-methyl ellagic acid, and cereotagloperoxide isolated from the ethyl acetate-soluble fraction	DPPH assay	184-186
<i>Cressa cretica</i>	<i>n</i> -Butanol, methanolic and ethyl acetate extracts	DPPH assay	187-189
<i>Crocus sativus</i>	petals, stigmas, entire flowers and crocin	DPPH, ABTS assay and <i>in vivo</i> antioxidant enzymes	190-198
<i>Crotalaria juncea</i>	Crude extracts	DPPH assay and <i>in vivo</i> antioxidant enzymes	199-200

<i>Cuminum cyminum</i>	Crude extracts and β -pinene, p-cymene, γ -terpinene, cuminaldehyde and cumin oils	DPPH assay and <i>in vivo</i> antioxidant enzymes	201-210
<i>Cupressus sempervirens</i>	The chloroform and methanol leaf extracts and essential oils	DPPH, ABTS assay and <i>in vivo</i> antioxidant enzymes	211-217
<i>Cydonia oblonga</i>	Crude leaves, fruits, pulps, peels and seeds, phenolic extracts and essential oils	DPPH assay, β -carotene–linoleic acid bleaching method, and lipid peroxidation inhibition assay	218-225
<i>Cynodondactylon</i>	Ethyl acetate, hexane, ethyl acetate, and methanol leaves extracts	DPPH, superoxide anion radical scavenging, nitric oxide scavenging assay, ferrous chelating ability, hydroxyl radical scavenging assay, hydrogen peroxide scavenging activity and ABTS assay	226-230
<i>Cyperus rotundus</i>	Extracts by different extraction solvents	DPPH, superoxide anion radicals, hydroxyl radicals, nitric oxide radical, hydrogen peroxide, in addition to property of metal chelating and reducing power.	231-233
<i>Dactylocteniumaegyptiacum</i>	Crude extract	DPPH method	234-235
<i>Dalbergia sisso</i>	ethanol extract of the bark; aqueous and methanol extracts of the stem bark	lipid peroxidation inhibitory (LPO), DPPH method	236-238
<i>Datura fastuosa</i>	different solvent extracts from the leaves	DPPH method, hydroxyl radical scavenging activity, reducing power assay, and β – carotene bleaching activity	239-241
<i>Datura stramonium</i>	Crude extracts	DPPH method	242
<i>Daucus carota</i>	Dry matter	TBARS and DPPH methods	243-245
<i>Desmostachyabipinnata</i>	methanolic extract	DPPH, nitric oxide, hydrogen peroxide, hydroxyl radical scavenging activities and DNA damage protection.	246-248
<i>Dianthus caryophyllus</i>	volatile oil	DPPH assay	249-250
<i>Digitalis species</i>	alcoholic extract of <i>Digitalis purpurea</i>	DPPH and the total antioxidant capacity	251-253
<i>Dodonaeaviscosa</i>	Ethanol and Methanolic extract	DPPH assay	254-255
<i>Dolichos lablab</i>	methanol extracts,	DPPH assay	256-257
<i>Echinochloa crus-galli</i>	Seeds and aerial parts extracts	DPPH assay	258-260
<i>Echium italicum</i>	Ethanol extracts	DPPH, Fe^{2+} - chelating ability, total phenolic contents and total flavonoid contents methods.	261-262
<i>Ephedra species</i>	Methanolic extract of <i>Ephedra alata</i>	DPPH assay	263-264
<i>Equisetum arvense</i>	Aqueous and ethanol extract	ABTS and DPPH assays	265-268
<i>Erigeron canadensis</i>	Methanolic, hexane, chloroform, ethyl acetate, butanol and 70% ethanolic extract	DPPH and against nitrate and oxidative damage induced by $ONOO^-$	269-272
<i>Erodium cicutarium</i>	Methanolic, extract, tannin, gallic acid, (+)-catechin and vitamin C	DPPH assay	273-276
<i>Eryngium creticum</i>	Aqueous and ethanolic extracts from different parts (leaves, stems, roots, and the whole plant)	DPPH assay	277-279
<i>Eucalyptus Species grown in iraq</i>	The essential oil and the subfractions of methanol extract from leaves of <i>Eucalyptus largiflorens</i> . The leaves extract and essential oil of <i>Eucalyptus camaldulensis</i>	DPPH	280-284
<i>Eupatorium cannabinum</i>	The hydro-alcoholic extract and caffeoyl derivatives	DPPH assay	285-287
<i>Euphorbia hirta</i>	different parts (leaves, stems, flowers and roots) extracts of <i>Euphorbia hirta</i>	DPPH assay	288-290
<i>Euphorbia tinctoria</i>	Extracts of the leaves and stems	DPPH assay	291-294
<i>Fagopyrum esculentum</i>	Seed components, epicatechin, (+)-	DPPH assay	295-298

	catechin 7-O-β-D-glucopyranoside, (–)-epicatechin 3-O-p-hydroxybenzoate, and (–)-epicatechin 3-O-(3,4-di-O-methyl) gallate		
<i>Ficus carica</i>	Leaves extracts, Cyanidin-3-rhamnoglucoside and crude hot-water soluble polysaccharide	DPPH assay	299-303
<i>Ficus carica</i>	Crude extracts	DPPH assay	304-305
<i>Ficus religiosa</i>	aqueous and ethanolic bark extracts	inhibition of lipid peroxidation assay	306-307
<i>Foeniculum vulgare</i>	Crude seeds extracts and 3-caffeoylquinic acid, 4-caffeoylquinic acid, 1.5-O-dicaffeoylquinic acid, rosmarinic acid, eriodictyol-7-O-rutinoside, quercetin-3-O-galactoside, kaempferol-3-O-rutinoside, kaempferol-3-O-glucoside, hydroxycinnamic acid derivatives, flavonoid glycosides and flavonoid aglycones	DPPH, FRAP assays and total antioxidant capacity	308-312
<i>Fraxinus ornus</i>	Ethanolic extract of bark, as well as esculetin, esculin, fraxetin and fraxin	DPPH assays and inhibition of lipid peroxidation	313-315
<i>Fumaria officinalis</i>	Methanolic extract of aerial parts	Ferric reducing antioxidant power assay, cupric reducing antioxidant capacity assay, DPPH radical scavenging, β-caroten-linoleic acid assay, metal chelating capacity and inhibition of the organic compounds and the conjugated diene hydroperoxides arising from linoleic acid oxidation	316-319
<i>Fumaria parviflora</i>	Crude extract	In vivo, evaluation of SOD, glutathione Peroxidase and glutathione reductase	320-321
<i>Galium aparine</i>	Methanol, ethyl acetate and ethanol crude aerial parts extracts	DPPH assay, H ₂ O ₂ scavenging assay and metal ion chelating ability	322-326
<i>Galium verum</i>	Aerial parts extracts	DPPH and nitric oxide radical scavenging, reducing power and H ₂ O ₂ scavenging	327-331
<i>Geum urbanum</i>	Crude extracts of different parts	DPPH, FRAP assays and percentage inhibition of linoleic acid oxidation capacity	332-334
<i>Glycyrrhiza glabra</i>	Roots extracts and Chalcone derivative, a novel group of neolignan lipid esters, and seven known phenolic compounds (formononetin, glabridin, hemileiocarpin, hispaglabridin B, isoliquiritigenin, 4'-O-methylglabridin, and paratocarpin B)	DPPH assay	335-339
<i>Gnaphalium luteoalbum</i>	Methanol extracts	DPPH assay	340-341
<i>Gossypium Species</i>	Hydroalcoholic, aqueous and ethanolic leaves extracts and gossypol	Free radical scavenging activity, reducing power assay, DNA damage prevention	342-346
<i>Haplophyllum species</i>	Aerial parts extracts and essential oils	β-carotene bleaching test, the reducing power test and H ₂ O ₂ induced cell damage	347-350
<i>Hedera helix</i>	Ethyl acetate, methanol and dichloromethane stem extracts, methanol extract of leaves, methanolic extract of the whole plant and α-hederin, hederasaponin-C, and hederacolchisides-E and F.	DPPH assay	351-355

<i>Helianthus annuus</i>	Methanol and hexane extracts of seeds	Inhibition of peroxidation of lipids, hydroxyl radical formation and DPPH radical formation	356-359
<i>Helianthus tuberosus</i>	Leaves and tubers extracts	Total antioxidants, ABTS and CUPRAC methods	360-363
<i>Helicophyllum crassipes (Eminiumspiculatum)</i>	Aqueous extracts of the leaves	Lipid peroxidation (inhibition	364
<i>Heliotropium</i> Species	Leaves, flowers stems, and roots extracts	DPPH assay	365-367
<i>Herniaria glabra and Herniariahirsuta</i>	Methanol extracts	DPPH assay	368-369
<i>Hibiscus cannabinus</i>	Flowers extracts	DPPH free radical scavenging activity in vitro and their capacity to protect DNA from oxidative damage <i>in vivo</i> .	370-371
<i>Hibiscus rosa-sinensis</i>	Different leaves and flowers extracts	DPPH free radical scavenging activity and percentage inhibition of linoleic acid oxidation capacity	372-379
<i>Hibiscus sabdariffa</i>	Different calyx, petal, and leaf extracts	(DPPH) inhibition, lipid peroxidation inhibition and <i>in vivo</i> antioxidant status	380-391
<i>Hyoscyamus species</i>	Arial parts and leaves extracts	DPPH antiradical, nitric oxide scavenging and metal chelating activities	392-399
<i>Hypericum triquetrifolium</i>	aerial parts extracts	DPPH free radical scavenging, ferric Reducingand ferrous chelating,	400-404
<i>Inula graveolens (Syn: Dittrichia graveolens)</i>	Methanolic leaves extract, methanolic and acetone extract of aerial parts	DPPH assay and ferrous ion chelating	405-410
<i>Jasminum officinale</i>	Crude aqueous, methanolic and ethanolic leaves extracts	DPPH, NO, superoxide and ABTS radicals in addition to reducing power assessment.	411-415
<i>Jasminum sambac</i>	Methanol and ethanol extracts and essential oils	DPPH free radical scavenging, β -carotene-linoleic acid assays andhydrogen peroxide method.	416-420
<i>Juglans regia</i>	Water, chloroform, methanol, ethanol, ethyl acetate and N-butanol extracts of the leaves, water, hydroalcoholic, chloroform and petroleum ether of the bark and polyphenols	DPPH radical scavenging, reductive power tests, H ₂ O ₂ scavenging activity, inhibition of lipid peroxidation andtotal antioxidant capacity	421-430
<i>Juncus maritimus</i>	Rhizomes extracts	DPPH assay	431
<i>Juniperus communis</i>	Water, ethanol extracts and essential oils	DPPH, ABTS, hydroxyl radical (OH \bullet) scavenging and chelating capacity, superoxide radical (\bullet O ₂ ⁻) scavenging, xanthine oxidase inhibitory effects and hydrogen peroxide scavenging.	432-436
<i>Juniperus oxycedrus</i>	Aqueous, ethanolic, methanolic extracts and essential oils	DPPH, TEAC, and FRAP	437-441
<i>Jussiaea repens</i>	Crude leaves extracts	DPPH assay	442-445
<i>Kochia scoparia (Syn: Bassia scoparia)</i>	Aqueous and 50% ethanol fructus kochiae extracts	DPPH radical assay, Superoxide anion radical scavenging activity and Lipid peroxidation inhibition assay	446-447
<i>Lagerstroemia indica</i>	Flowers and leaves extracts	DPPH, ABTS and FRAP assays	448-451
<i>Lagerstroemia speciosa</i>	Leaves and barks extracts	DPPH assay	452-455
<i>Lallemantiaiberica</i>	Ethyl acetate and methanol leaves extracts and essential oils	DPPH and FRAP assays	456-458
<i>Lallemantiaroyleana</i>	Seeds different extracts	DPPH and FRAP assays	459-460
<i>Lantana camara</i>	Extracts of leaves and Stem, lantadene A, oleanolic acid and lantanilic acid	DPPH, the total phenolic and flavonoids contents and total antioxidant activity	461-466
<i>Lathyrus sativus</i>	Crude extracts	DPPH scavenging activity, reducing power, β -carotene bleaching inhibition and TBARS formation inhibition	467-471
<i>Lawsoniainermis</i>	Leaves methanol, petroleum ether and	DPPH and ABTS assays	472-478

	ethyl Acetate extracts		
<i>Lemna minor</i>	lyophilized water and ethanol extracts	H ₂ O ₂ scavenging activity, Ferrous ion chelating activity and Superoxide scavenging activity	479-480
<i>Leonticeleontopetalum</i>	Lupanine, a quinolizidine alkaloid isolated from the tubers	ABTS, FRAP and reduction power	481-483
<i>Lepidium sativum</i>	Methanolic, ethyl acetate and ethanolic extracts	DPPH, ABTS, superoxide scavenging activity and metal chelating property	484-489
<i>Lippianodiflora</i>	Defatted methanolic extract of aerial parts, methanol and ethyl acetate extract of leaf and stem	DPPH assay and inhibition of lipid peroxidation	490-494
<i>Luffa acutangula</i>	Ethyl acetate and ethanol extracts of dried leaves, peels and seeds extracts	DPPH, ABTS, superoxides radical, reducing power and phosphomolybdenum assay.	495-499
<i>Luffa cylindriica</i>	Chloroform, n-hexane, ethyl acetate, ethanol and methanol extracts of leaves	ferric thiocyanate test, thiobarbituric acid test, ferric reducing antioxidant power and DPPH free radicals scavenging test	500-504
<i>Lycopus europaeus</i>	Leaves waterand hydroalcoholicextracts	DPPH and ABTS assays	505-507
<i>Lythrumsalicaria</i>	Methanolic and ethanolic extracts of the aerial parts	DPPH, nitric oxide and hydrogen peroxide scavenging activities	508-513
<i>Malva neglecta</i>	Leaves, stems, flowers and roots extracts	DPPH, FRAP and ORAC assays.	514-518
<i>Mangifera indica</i>	Aqueous-methanolic extracts of pulp, peel and seed kernels, many crude extracts, fractions (ethyl acetate and n-butanol) and many separated compounds	DPPH, FRAPAssays, and inhibition of lipid peroxidation	519-527
<i>Marrubium vulgare</i>	Methanolic extract, essential oils and flavonoids (acacetin, apigenin, and acacetin-7-rhamnoside).	DPPH and nitric oxide antioxidant activity and LPO inhibition	528-532
<i>Matricaria chamomilla</i>	Crude extracts of different parts and polyphenolic-polysaccharide conjugates	Various antioxidant assays: DPPH, ABTS, linoleic acid emulsion, ferric ions reducing antioxidant power, ferrous ions chelating capacity, superoxide radical scavenging activity assays	533-537
<i>Medicago sativa</i>	Flowers extracts, raw seeds and germinated seeds	DPPH and FRAP assays	538-542
<i>Melilotus officinalis</i>	Hexane, 96 and 50% ethanol extract	DPPH assay and lipid peroxidation inhibition	543-546
<i>Melissa officinalis</i>	Hydroalcoholic extract and infusions	DPPH radical scavenging model and CUPRAC method	547-549
<i>Mirabilis jalapa</i>	Acetone, ethyl acetate, petroleum ether, methanol and ethanol extracts of leaves	The total antioxidant capacity, FRAP and DPPH tests.	550-553
<i>Musa paradisiaca</i>	The aqueous and ethanolic extracts	DPPH assay	554-556
<i>Narcissus tazetta</i>	Bulb and 11 compounds [tazettones C-G, (2S)-3',4'-dihydroxy-7-methoxyflavan, (2S)-3',7-dihydroxy-4'-methoxy8-methylflavan, (2S)-liquiritigenin 7-methyl ether, 8-methylnaringenin, farrerol, and cyrtominetin], isolated from the bulbs.	DPPH and FRAP assays.	557-559
<i>Nasturtium officinale</i>	Ethanolic extract, phenolic, flavonoid, and anthocyanin contents	DPPH, ABTS and Inhibition of lipid peroxidation assays	560-563
<i>Nerium oleander</i>	Essential oil and different extracts (water, methanol, water: methanol and acetone).	DPPH assay; β -Carotene/linoleic acid a bleaching assay and ferric reducing power assay.	564-567

<i>Nicotiana tabacum</i>	The extracts of the stem, flavonoids and polysaccharides.	DPPH, ABTS, and reducing power tests.	568-572
<i>Ocimum basilicum</i>	Flavonoids, polyphenols, essential oil and crude leaf extracts.	DPPH assay	573-578
<i>Olea europaea</i>	Ethanol, hydro-alcoholic and aqueous extracts of leaves, and phenolic compounds	DPPH, ABTS, total antioxidant capacity (TAC), xanthine oxidase (XO) inhibitory assays.	579-584
<i>Ononis spinosa</i>	Crude extracts and fractions	DPPH and ABTS assays.	585-588
<i>Onopordonacanthium</i>	Butanolic, ethanol, and acetone extracts from flowers and leaves	DPPH assay.	589-592
<i>Orchis masscula</i>	Polysaccharide and crude extracts	DPPH method and total antioxidant effect.	593-595
<i>Origanum vulgare</i>	The essential oil and aqueous, ethanolic and methanolic extracts	DPPH and ABTS assays.	596-600
<i>Oxalis corniculata</i>	Flavonoids, crude methanolic and ethanolic leaves extracts.	DPPH, nitric oxide radical scavenging activity and against MPTP induced oxidative stress.	601-605
<i>Phoenix dactylifera</i>	The fruit extracts and polyphenol, flavonoid, anthocyanin contents.	FRAP and DPPH assays.	606-612
<i>Prunus armeniacae</i>	Fruits, water and methanol extracts of sweet kernels	FRAP	613-614
<i>Ranunculus sceleratus</i>	Crude extracts of <i>Ranunculus sceleratus</i>	DPPH, FRAP, TEAC, CUPRAC and SNP (silver nanoparticle assay)	615-616
<i>Ranunculus arvensis</i>	Crude extracts of <i>Ranunculus arvensis</i>	DPPH	617-618
<i>Reseda Species</i>	Methanol extracts of the flowers and leaves of <i>Reseda lutea</i>	On the activity of catalase, glutathione-S-transferase, and glutathione peroxidase	619-620

Plant antioxidants are mainly polyphenols (phenolic acids, flavonoids, anthocyanins, lignans and stilbenes), carotenoids (xanthophylls and carotenes) and vitamins (vitamin E and C) ¹⁴⁻⁵¹.

The antioxidant effects of medicinal plants are determined by many chemical-based assays. Among these assays, some are based on the ability to scavenge stable free radicals, such as Trolox equivalence antioxidant capacity (TEAC), DPPH assay and, Folin–Ciocalteu reagent assay, and some assays are based on the ability to reduce metal ions, such as ferric ion reducing antioxidant power (FRAP), and cupric reducing antioxidant capacity (CUPRAC). Meanwhile, HAT-based assays detect the ability of an antioxidant to quench free radicals by hydrogen donation, which is more relevant to the radical chain-breaking antioxidant capacity. HAT-based assays include oxygen radical absorbance capacity (ORAC), total radical trapping antioxidant parameter (TRAP), and inhibiting the oxidation of low-density lipoprotein (LDL) ¹⁶.

This review highlighted the natural sources of antioxidants to be used as beneficial supplements in prevention of many diseases.

CONCLUSION:

Antioxidants from plant sources are biologically active compounds that have medicinal and therapeutic values as alternatives to their synthetic analogues, due to their

stability, cheapness, and lack of undesirable effects associated with their use. This review presented plant sources of antioxidants to encourage further studies on their effectiveness when used medicinally, their stability, their pharmacokinetics, and the possibility of side effects resulting from them.

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